Supplementary Material

Synthesis of new heterocycle-based selenoamides as potent cytotoxic agents

J. G. García-López,^a A.I. Gutiérrez-Hernández,^b R. A. Toscano,^b M. T. Ramírez-Apan,^b J. A. Terrón,^a M. C. Ortega-Alfaro,^c and J. G. López-Cortés^{*b}

^aDepartamento de Farmacología, Centro de Investigaciones y Estudios Avanzados CINVESTAV-IPN, Av. IPN, Gustavo A. Madero, CP 07360, Cd. Mx. México ^bInstituto de Química, Universidad Nacional Autónoma de México, Circuito Exterior, Ciudad Universitaria, Coyoacán, CP 04510 Cd. Mx. Mexico ^cInstituto de Ciencias Nucleares, Universidad Nacional Autónoma de México, Circuito Exterior, Ciudad Universitaria, Coyoacán, C.P. 04510 Cd. Mx. Mexico Email: jqlcvdw@unam.mx

Table of Contents

1.	Materials and instruments	S2
2.	Synthesis of carbene complexes 1(a-h)	S2
3.	NMR and HR-M spectra of 1c	S3
4.	Synthesis of aminocarbene complexes 2(a-h)	S5
5.	NMR and HR-M spectra of 2(c-g)	S7
6.	Synthesis of selenoamides 3(a-h)	.S16
7.	NMR and HR-M spectra of 3(a-g)	.S19
8.	X-Ray diffraction analyses of 3f	. S31
9.	Biological Studies	S32
10.	References	S33

1. - Materials and instruments

THF and diethyl ether were distilled from sodium/benzophenone under a nitrogen atmosphere. All reagents and solvents were obtained from commercial suppliers and used without further purification. All compounds were characterized by IR spectra, recorded on a Bruker Tensor 27 spectrophotometer, by KBr or film techniques, and all data are expressed in wave numbers (cm⁻¹). Melting points were obtained on a Melt-Temp II apparatus and are uncorrected. NMR spectra were measured with a JEOL Eclipse +300 and Bruker Avance III 300, using CDCl₃ and CD₃CN as solvents. Chemical shifts are in ppm (δ), relative to TMS. The MS-EI were obtained on a JEOL JMS-AX505 HA using 70 eV as ionization energy and for MS-FAB a JEOL JMS-SX 102A using nitrobenzyl alcohol and polyethylene glycol as matrix. All tested compounds synthesized are more than 95 % pure, analyzed using HPLC HP 1100 with diode-array detector.

2. - Synthesis of Fischer ethoxyarylcarbene chromium 0 complexes 1(a-h)

The preparation of Fischer-type carbene complexes was carried out using a slightly modification of the methodology previously described elsewhere.¹ To a solution of the corresponding aryl substrate (8 mmol) in 10 mL of anhydrous THF under argon atmosphere was added at 0°C a solution of *n*BuLi (8.2 mmol). The reaction mixture was stirred at room temperature for 20 to 60 min. and then transferred by canula to a suspension of Cr(CO)₆ (1.74 g, 8 mmol) in THF (20 mL). The mixture was then stirred for the time specified in Table 1 of main document, at room temperature. The solvent was removed under vacuum, and then triethyloxonium tetrafluoroborate (2 g, 10 mmol) on ice/water was added. The organic phase was washed with saturated solution of NaHCO₃ and then with brine. The organic phase was dried with anhydrous sodium sulfate and the solvent was evaporated under vacuum. The mixture was purified by chromatography on silica gel or alumina using hexane as eluent. Fischer carbene complexes **1a**, **1b**, **1d**, **1e**, **1f**, **1g** and **1h** are already known and their spectroscopic data match well with literature.¹⁻³

[(ethoxy)(benzo[*d*][1,3]dioxol-5-yl)methylidene]pentacarbonyl chromium (0) (1c): Yield: 60 %; mp 66 – 70 (d) °C; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 7.36 (d, *J*= 7.2 Hz, 1 H; C₆H₃), 7.03 (s, 1 H; C₆H₃), 6.86 (d, *J*= 7.8 Hz, 1 H; C₆H₃), 6.05 (s, 2 H; OCH₂O), 5.12 (d, 2 H; OCH₂), 1.70 (s, 3H, CH₃) ppm. ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 339.8 (C=Cr), 223.8 (CrCO_{*ax*}), 216.7 (CrCO_{*eq*}), 150.9 (C, C₆H₃), 147.9 (C, C₆H₃), 147.6 (C, C₆H₃), 124.5 (CH, C₆H₃), 107.6 (CH, C₆H₃), 105.2 (CH, C₆H₃), 101.9 (OCH₂O), 76.6 (OCH₂), 15.2 (CH₃) ppm. IR (KBr) (cm⁻¹): 2057, 1905 (CrCO). MS (EI, 70 eV) *m/z* (%): 370 (8) [*M*⁺], 342 (27) [*M*⁺-CO], 314 (47) [*M*⁺-2CO], 286 (38) [*M*⁺-3CO], 258 (92) [*M*⁺-4CO], 230 (68) [*M*⁺-5CO]. HRMS (FAB⁺): *m/z*: calcd. for C₁₅H₁₀CrO₈: 369.9781 [*M*⁺]; found: 369.9768.

3. - NMR spectra of 1c



Figure S2. ¹³C NMR spectrum of **1c**

Page: 1 [Elemental Composition] Date :13-Mar-2013 15:42 Data : Dr-Jose-G-Lopez113 Sample: 657 OOCrOEt Note : Luis-Velasco Inlet : Direct Ion Mode : FAB+ Scan#: (1,8) RT : 0.66 min Elements : C 30/0, H 49/0, O 9/0, Cr 2/0 Mass Tolerance : 100ppm, 2mmu if m/z >2 Int% Observed m/z 369.9768 12.7 Estimated m/z Error [ppm] U.S.C H O Cr 13 15 10 8 1 369.9781 -0.8

Figure S3. HR-MS of compound 1c

4. - Synthesis of Fischer Aminocarbenes chromium 0 complexes 2(a-h) (Route A)

To a solution of Fischer ethoxycarbene complex (1) (2.3 mmol) in 20 mL of anhydrous diethyl ether under nitrogen atmosphere was added 4.9 mmol of ethanolamine. The reaction mixture was stirred at room temperature for 5 to 20 min. and then diluted with 20 mL of water. The organic phase was separated and dried with anhydrous Na₂SO₄, and the solvent was evaporated in vacuum. The crude product was purified by flash column chromatography using alumina and Hexane-AcOEt mixture (95:5) as eluent. Compounds **2a**, **2b** and **2h** are known and their spectroscopic data match well with already described in the literature.¹⁻³

[(benzo[*d***][1,3]dioxo-5-yl)(2-hydroxyethylamino)methylidene]pentacarbonyl chromium (0) (2c)** (Unstable in solution) :Yield: 81 %; Yellow oil; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 9.51 (s, 1 H; NH_{*E*}), 9.06 (s, 1 H; NH_{*Z*}), 6.83 (s, 1 H; C₆H₃), 6.34 – 6.26 (m, 2H; C₆H₃), 5.99 (s, 2H; OCH₂O), 3.76 (s, 2 H; OCH₂), 3.39 (s, 2 H, NCH₂), 1.25 (s, 1 H; OH) ppm. IR (Film), cm⁻¹: 3410 (OH), 3352 (NH), 2054, 1974, 1911 (Cr-CO). MS (FAB⁺) *m/z* (%): 385 (12) [*M*⁺], 357 (16) [*M*⁺-CO], 329 (15) [*M*⁺-2CO], 301 (21) [*M*⁺-3CO], 273 (28) [*M*⁺-4CO], 245 (72) [*M*⁺-5CO]; HRMS (FAB⁺) *m/z*: calcd. for C₁₅H₁₁CrNO₈: 384.9890 [*M*⁺]; found: 384.9888.

[(*N*-methylpyrrol-2-yl)(2-hydroxyethylamino)methylidene]pentacarbonyl chromium (0) (2d): Yield: 90%; Yellow oil; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 9.40 (s, 1 H; NH_{*E*}), 8.62 (s, 1 H; NH_{*Z*}), 6.61 and 6.55 (s, 1 H; C₄H₃N), 6.17 (s, H; C₄H₃N), 5.97 (s, H; C₄H₃N), 3.54-3.38 (m, 7 H; OCH₂, NCH₂, NCH₃) 1.25 (s, 1 H; OH) ppm. ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 277.2 and 265.4 (CCr, Isomers *E* and *Z*), 223.5 (CO_{*ax*} Isomer *E*), 217.7 (CO_{*eq*} Isomers *Z*), 217.2 (CO_{*eq*} Isomer *E*), 211.5 (CO_{*eq*} Isomer *Z*), 163.3 (C, C₄H₃N), 145.1 (CH, C₄H₃N), 123.2 (CH, C₄H₃N), 109.0 and 106.1 (CH, C₄H₃N, Isomers *E* and *Z*), 62.9 and 60.9 (OCH₂, Isomers *E* and *Z*), 54.4 and 52.6 (NCH₂, Isomers *E* and *Z*), 52.6 ppm (NCH₃). IR (Film) cm⁻¹: 3609 (OH), 3373 (NH), 1998, 1870 (CrCO). MS (FAB⁺) *m/z* (%): 345 (80) [*M*⁺+1], 317 (12) [*M*⁺+1-CO], 289 (65) [*M*⁺+1-2CO], 261 (100) [*M*⁺+1-3CO], 233 (57) [*M*⁺+1-4CO], 204 (14) [*M*⁺+1-5CO]; HRMS (FAB⁺) *m/z*: calcd. for C₁₃H₁₂CrN₂O₆: 344.0100 [*M*⁺]; found: 344.0110.

[(*N*-methylindol-2-yl)(2-hydroxyethylamino)methyliden]pentacarbonyl chromium (0) (2e): Yield: 92 %; Yellow oil; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 9.92 (s, 1 H; NH), 7.59 (d, *J*= 7.5 Hz, 1 H; C₈H₅N), 7.33 – 7.12 (m, 3 H; C₈H₅N), 6.18 (s, 1 H; C₈H₅N), 3.72 - 360 (m, 7 H; OCH₂, NCH₂, NCH₃), 3.34 (s, 1 H; OH) ppm. ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 279.0 (CCr), 223.1 (CrCO_{ax}), 216.8 (CrCO_{eq}), 145.0 (C, C₈H₅N), 137.0 (C, C₈H₅N), 128.1 (C, C₈H₅N), 122.1, (CH, C₈H₅N), 121.0 (CH, C₈H₅N), 120.4 (CH, C₈H₅N), 109.4 (CH, C₈H₅N), 96.0 (CH, C₈H₅N), 60.6 (OCH₂), 50.3 (NCH₂), 30.8 (NCH₃) ppm. IR (Film) cm⁻¹: 3349 (OH) and (NH), 2054, 1976, 1912 (CrCO). MS (FAB⁺) *m/z* (%): 395

(11) [*M*⁺+1], 366 (62) [*M*⁺-CO], 338 (18) [*M*⁺-2CO], 310 (11) [*M*⁺-3CO], 282 (100) [*M*⁺-4CO], 254 (84) [*M*⁺-5CO]. HRMS (FAB⁺) *m/z*: calcd. for C₁₇H₁₄CrN₂O₆: 394.0257 [*M*⁺]; found: 394.0260.

[(thien-2-yl)(2-hydroxyethylamino)methyliden]pentacarbonyl chromium (0) (2f): Yield: 95%; m.p. 105 -107°C (d); ¹H NMR (300 MHz, CDCl₃, TMS): δ = 9.45 (s, 1 H; NH_{*E*}), 9.13 (s, 1 H; NH_{*Z*}), 7.47 and 7.41 (m, 2 H; C₄H₃S, Isomers *E* and *Z*), 7.11 and 7.06 (m, 2 H; C₄H₃S, Isomers *E* and *Z*), 6.84 (s, 1H; C₄H₃S) 4.27 and 4.09 (d, 4 H; OCH₂, Isomers *E* and *Z*), 3.86 and 3.66 (d, 4 H; NCH₂, Isomers *E* and *Z*), 1.99 (s, 1 H; OH) ppm. ¹³C NMR (75 MHz, CDC₁₃, TMS): δ = 272.9 and 261.2 (CCr, Isomers *E* and *Z*), 223.2 (CO_{*ax*}, Isomers *E* and *Z*), 217.5 and 217.1 (CO_{*eq*}, Isomers *E* and *Z*), 155.7 and 149.01(C, C₄H₃S, Isomers *E* and *Z*), 129.3 and 128.2 (CH, C₄H₃S, Isomers *E* and *Z*), 127.6 and 127.3 (CH, C₄H₃S, Isomers *E* and *Z*), 126.7 and 122.6 (CH, C₄H₃S, Isomers *E* and *Z*), 61.0 and 60.9 (OCH₂), 54.6 and 52.3 ppm (NCH₂, Isomers *E* and *Z*) ppm. IR (KBr) cm⁻¹: 3581 (OH), 3308 (NH), 2053, 1869 (Cr-CO). MS (EI, 70 eV) *m/z* (%): 347 (14.96) [*M*⁺], 319 (7.04) [*M*⁺-CO], 291 (8.16) [*M*⁺-2CO], 263 (20.41) [*M*⁺-3CO], 235 (36.73) [*M*⁺⁻4CO], 207 (13.6) [*M*⁺⁻5CO]. HRMS (FAB⁺) *m/z*: calcd for C₁₂H₉CrNO₆S: 346.9556 [*M*⁺]; found: 346.9560.

[(furan-2-yl)(2-hydroxyethylamino)methyliden]pentacarbonyl chromium (0) (2g): Yield: 96%; m.p. 85-86 °C; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 9.73 (s, 1 H; NH_{*E*}), 8.89 (s, 1 H; NH_{*Z*}), 7.67 and 6.48 (s, 2H; C₄H₃O, Isomers *E* and *Z*), 7.19 (s, 1 H; C₄H₃O), 6.61 (s, 2 H; C₄H₃O, Jomers *E* and *Z*), 4.26 - 3.92 (m, 8 H; OCH₂, NH₂, Jomers *E* and *Z*), 1.88 (s, 1 H; OH) ppm. ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 244.2 (CCr), 222.7 (CO_{ax}), 218.0 and 217.8 (CO_{eq} Isomers *E* and *Z*), 145.8 and 144.2 (C, C₄H₃O, Isomers *E* and *Z*), 125.1 and 123.4 (CH, C₄H₃O, Isomers *E* and *Z*), 113.4 and 112.9 (CH, C₄H₃O, Isomers *E* and *Z*), 61.0 and 60.5 (OCH₂, Isomers *E* and *Z*), 54.1 (NCH₂) ppm. IR (KBr) cm⁻¹: 3581 (OH), 3308 (NH), 2053, 1869 (Cr-CO). MS (EI, 70 eV) *m/z* (%): 331 (15) [*M*⁺], 303 (7) [*M*⁺-CO], 275 (8) [*M*⁺-2CO], 247 (20) [*M*⁺-3CO], 219 (37) [*M*⁺-4CO], 191 (14) [*M*⁺-5CO]. HRMS (FAB+) *m/z*: calcd for C₁₂H₉CrNO₇: 330.9784 [*M*⁺]; found: 330.9792.

5. - NMR and HR-MS spectra of 2c-g



Figure S4. ¹H NMR spectrum of **2c** (*E*/*Z* ratio 72:28)

[Elemental Composition] Data : Dr-Jose-G-Lopez038 Sample: 579 OOCrNEtOH Note : Luis-Velasco	Page: 1 Date :16-Mar-2013 19:28
Inlet : Direct RT : 0.62 min Elements : C 30/0, H 49/0, O 9/0, Mass Tolerance : 100ppm, 2mmu	Ion Mode : FAB+ Scan#: (1,6) N 4/1, Cr 2/0 if m/z >2
Observed m/z Int% 384.9888 11.2 Estimated m/z Error [ppm] U.S. 384.9890 -0.5 13	C H O N Cr 15 11 8 1 1

Figure S5. HR-MS of compound 2c





Figure S7. ¹³C NMR spectrum of **2d**

[Elemental Composition] Data : Dr-Jose-G-Lopez268 Sample: 1433 PirrCrN Note : Luis-Velasco	Page: 1 Date :09-Aug-2017 11:37
Inlet : Direct	Ion Mode : FAB+
RT : 0.62 min	Scan#: (1,6)
Elements : C 30/0, H 49/0, O 9/0,	N $4/1$, Cr $2/0$
Mass Tolerance : 100ppm, 2mmu	if $m/z > 2$
Observed m/z Int%	
344.0110 75.1	
Estimated m/z Error [ppm] U.S.	C H O N Cr
344.0100 2.9 11	13 12 6 2 1

Figure S8. HR-MS of compound 2d



Figure S10. ¹³C NMR spectrum of **2e**

Page: 1 [Elemental Composition] Data : Dr-Jose-G-Lopez093 Date :30-Jan-2013 18:31 Sample: 186 MeIndCrNEtOH Note : Luis-Velasco Inlet : Direct Ion Mode : FAB+ Scan#: (4,8) RT : 0.96 min Elements : C 30/0, H 49/0, O 9/0, N 4/1, Cr 2/0 Mass Tolerance : 100ppm, 2mmu if m/z >2 Observed m/z Int% 394.0260 58.2 CrEstimated m/z Error [ppm] U.S.C H 0 N 14 17 14 6 2 1 394.0257 -0.7

Figure S11. HR-MS of compound 2e



Figure S13. ¹³C NMR spectrum of **2f**

[Elemental Composition] Page: 1 Date :30-Jan-2013 18:10 Data : Dr-Jose-G-Lopez093 Sample: 186 SCrNEtOH Note : Luis-Velasco Inlet : Direct Ion Mode : FAB+ RT : 0.85 min Scan#: (3,8) Elements : C 30/0, H 49/0, O 9/0, N 4/1, S 2/0, Cr 2/0 Mass Tolerance : 100ppm, 2mmu if m/z > 2Observed m/z Int% 346.9560 22.5 Estimated m/z Error [ppm] U.S.C H O N S Cr 346.9556 1.1 11 12 9 6 1 1 1

Figure S14. HR-MS of compound 2f



Figure S15. ¹H NMR spectrum of **2g** (*E*/*Z* ratio 55:45)



Figure S16. ¹³C NMR spectrum of **2g**

[Elemental Composition] Page: 1 Date :01-Feb-2013 12:19 Data : Dr-Jose-G-Lopez103 Sample: 279 OCrNOH Note : Luis-Velasco Inlet : Direct Ion Mode : FAB+ RT : 0.93 min Scan#: (2,7) Elements : C 30/0, H 49/0, O 9/0, N 4/1, Cr 2/0 Mass Tolerance : 100ppm, 2mmu if m/z >2 Observed m/z Int% 330.9792 17.7 Estimated m/z Error [ppm] U.S.C H O N Cr 11 12 9 7 1 330.9784 2.4 1

Figure S17. HR-MS of compound 2g

6. - Synthesis of selenoamides 3(a-f) (Route A)

Table S1. Synthesis of arylselenoamides 3(a-f)



Route B: Sequence	aminolysis-demetalation	one-pot reaction
	,	

	Ar		Time (min)	Route A		Route B	
Entry		Compound	Demetalation step	Yield (%) ^a	Global Yield (%)ª	Yield (%) ^b	Global Yield (%) ^b
1		За	25	94	70	93	74
2	ې ۲	3b	20	96	78	95	80
3	o o	3c	35	91	44	90	64
4	N,	3d	45	83	46	90	64
5	N Start	Зе	45	72	63	80	75
6	S S	3f	35	90	60	91	76
7		3g	45	90	69	92	74
8	Fc	3h	15	91	77	95	81

For ^aRoute A and ^bRoute B, the global yield was determined from the corresponding aryl substrate used as starting material.

Preparation of selenating agent: To a solution of 0.01 mol of NaBH₄ in 10 mL of ethanol was added 0.01 mol of powdered selenium, and the mixture was vigorously stirred at room temperature for 30 min under nitrogen atmosphere. The selenating agent was then added to a solution (0.001 mol) of the corresponding aminocarbene complex in 5 mL of ethanol, under nitrogen atmosphere; the reaction was monitored by TLC on silica-gel. After

the reaction was completed, the solvent was evaporated under vacuum, the residual mixture was dissolved in distilled water and the product was extracted with CH_2Cl_2 and then dried with anhydrous Na_2SO_4 . After the evaporation of the solvent, the resultant mixture was purified by silica-gel column using a mixture Hexane: CH_2Cl_2 (1:1) as eluent. Selenoamides **3a** and **3f** are known and their spectroscopic data match well with literature.³

N-(2-hydroxyethyl)-4-methoxybenzenecarboselenoamide (3b). Yield: 97%; m.p. 68 - 73°C; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 8.51 (s, 1 H; NH), 7.79 (d, *J*= 8.4 Hz, 2 H; C₆H₄), 6.86 (d, *J*= 8.4 Hz, 2 H; C₆H₄), 4.07 – 4.00 (m, 4 H; OCH₂, NCH₂), 3.83 (s, 3 H; OCH₃), 2.17 (s, 1 H; OH) ppm. ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 203.0 (C=Se), 162.3 (C, C₆H₄), 137.0 (C, C₆H₄), 128.7 (2 CH, C₆H₄), 113.7 (2CH, C₆H₄), 58.9 (OCH₂), 55.5 (OCH₃), 49.7 (NCH₂) ppm. IR (KBr) cm⁻¹: 3377 (NH), 1607 (C=Se); MS (EI, 70 eV) *m/z* (%): 259 (7) [*M*⁺], 177 (97) [*M*⁺-H₂Se], 134 (15) [ArCNH⁺]; HRMS (FAB⁺): *m/z*: calcd. for C₁₀H₁₃NO₂Se: 259.0112 [*M*⁺]; found: 259.0100.

N-(2-hydroxyethyl)benzo[*d*][1,3]dioxole-5-carboselenoamide (3c). Yield: 90%; m.p. 254 - 256°C; ¹H NMR (300 MHz, CD₃CN, TMS): δ = 9.05 (s, 1 H; NH), 7.33 - 7.31 (m, 2 H; C₆H₃), 6.80 (d, 1 H; C₆H₃), 6.00 (s, 2 H; OCH₂O), 3.88-3.82 (m, 4 H; OCH₂, NCH₂), 3.05 (s, 1 H; OH) ppm. ¹³C NMR (75 MHz, CD₃CN, TMS): δ = 202.0 (C=Se), 149.8 (C, C₆H₃), 147.3 (C, C₆H₃), 138.6 (C, C₆H₃), 121.2 (CH, C₆H₃), 107.6 (CH, C₆H₃), 107.0 (CH, C₆H₃), 101.9 (CH₂, OCH₂O) 58.4 (OCH₂), 52.0 (NCH₂) ppm. IR (KBr) cm⁻¹: 3357 (NH), 1607 (C=Se). MS (EI, 70 eV) *m/z* (%): 273 (40) [*M*⁺], 191 (14) [*M*⁺-H₂Se], 148 (100) [ArCNH⁺]; HRMS (FAB⁺): *m/z*: calcd for C₁₀H₁₂NO₃Se: 273.9982 [*M*⁺+1]; found: 273.9977.

N-(2-hydroxyethyl)-1-methyl-1*H*-pyrrole-2-carboselenoamide (3d). Yield: 93%; m.p. 79 - 80°C; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 7.86 (s, 1 H; NH), 6.78 (s, 1 H; C₄H₃N), 6.48 - 6.47 (m, 1 H; C₄H₃N), 6.08 - 6.06 (m, 1 H; C₄H₃N), 3.99 (s, 3 H; NCH₃), 3.93 - 3.90 (m, 4 H; NCH₂CH₂), 2.45 (s, 1 H; OH) ppm. ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 190.4 (C=Se), 137.2 (C, C₄H₃N), 130.5 (CH, C₄H₃N), 110.2, (CH, C₄H₃N), 107.7 (CH, C₄H₃N), 60.3 (OCH₂), 50.2 (NCH₂), 37.3 (NCH₃) ppm. IR (KBr) cm⁻¹: 3351 (NH), 1652 (C=Se). MS (EI, 70 eV) *m/z* (%): 232 (8) [*M*⁺], 149 (100) [*M*⁺-H₂Se], 107 (24) [*M*⁺- ArCNH⁺]; HRMS (FAB⁺): *m/z*: calcd. for C₈H₁₂N₂OSe: 232.0115 [*M*⁺]; found: 232.0122.

N-(2-hydroxyethyl)-1-methyl-1*H*-indole-2-carboselenoamide (3e). Yield: 72 %; m.p.70 - 71°C; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 8.63 (s, 1 H; NH), 7.63 (d, 1 H; C₈H₅N), 7.34 (s, 2 H; C₈H₅N), 7.14 (s, 1 H; C₈H₅N), 6.73 (s, 1 H; C₈H₅N), 4.02 – 4.01 (m, 7H; OCH₂, NCH₂, NCH₃) 1.65 ppm (s, 1 H; OH). ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 192.5 (C=Se), 143.3 (C, C₈H₅N), 140.3 (C, C₈H₅N), 126.3, (C, C₈H₅N), 124.2 (CH, C₈H₅N), 121.9 (CH, C₈H₅N), 121.0

(CH, C₈H₅N), 110.4 (CH, C₈H₅N), 101.7 (CH, C₈H₅N), 60.4 (OCH₂), 50.6 (NCH₂), 32.4 ppm (NCH₃). IR (KBr): *v* = 3421 (OH), 3150 (NH), 1644 cm⁻¹ (C=Se). MS (EI, 70 eV) *m/z* (%): 282 (100) [*M*⁺], 201 (29) [*M*⁺-H₂Se], 157 (79) [ArCNH⁺]. HRMS (FAB⁺) *m/z*: calcd. for C₁₂H₁₄N₂OSe: 282.0271 [*M*⁺]; found: 282.0285.

N-(2-hydroxyethyl)thiophene-2-carboselenoamide (3f). Yield: 75%; m.p. 118-120°C; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 8.42 (s, 1 H; NH), 7.60 (d, *J*= 6.0 Hz, 1 H; C₄H₃S), 7.52 (d, *J*= 3.0 Hz, 1 H; C₄H₃S), 7.13 (d, *J*= 3.0 Hz, 1 H; C₄H₃S), 4.08 - 4.04 (m, 4 H; OCH₂, NCH₂), 1.87 ppm (s, 1 H; OH). ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 192.2 (C=Se), 149.8 (C, C₄H₃S), 133.2 (CH, C₆H₄), 128.2 (CH, C₄H₃S), 124.4 (CH, C₄H₃S), 60.4 (OCH₂), 51.2 ppm (NCH₂). IR (KBr): v = 3370 (NH), 1658 cm⁻¹ (C=Se). MS (EI, 70 eV) *m/z* (%): 235 (100) [*M*⁺], 154 (32) [*M*⁺-H₂Se], 110 (89) [ArCNH⁺]. HRMS (FAB⁺) *m/z*: calcd. for C₇H₉NOSSe: 234.9570 [*M*⁺]; found: 234.9582.

N-(2-hydroxyethyl)furan-2-carboselenoamide (3g). Yield: 97%; m.p. 74-76°C; ¹H NMR (300 MHz, CDCl₃, TMS): δ = 8.90 (s, 1 H; NH), 7.55 – 7.50 (m, 2 H; C₄H₃O), 6.47 (s, 1 H; C₄H₃O), 4.06 - 3.98 (m, 4 H; OCH₂, NCH₂), 2.51 ppm (s, 1 H; OH). ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 185.1 (C=Se), 155.1 (C, C₄H₃O), 144.4 (CH, C₄H₃O), 120.8 (CH, C₄H₃O), 113.7 (CH, C₄H₃O), 60.4 (CH, OCH₂) 50.0 ppm (NCH₂). IR (KBr): v = 3330 (NH), 1532 cm⁻¹ (C=Se). MS (EI, 70 eV) *m/z* (%): 219 (100) [M+], 138 (18) [M+-H2Se], 94 (83) [ArCNH⁺]. HRMS (FAB+) *m/z*: calcd. for C₇H₉NO₂Se: 218.9799 [M+]; found: 218.9785.

7. - NMR and HR-MS spectra of 3b-g



[Elemental Composition] Page: 1 Data : Dr-Jose-G-Lopez037 Date :28-Jul-2016 11:39 Sample: 951 AIGH-5b Note : Luis-Velasco Inlet : Direct Ion Mode : FAB+ RT : 0.32 min Scan#: (8,10) Elements : C 30/0, H 49/0, O 3/0, N 3/0, Se 1/0 Mass Tolerance : 100ppm, 2mmu if m/z >2 Observed m/z Int% 259.0100 70.1 Estimated m/z Error [ppm] U.S.C H O N 259.0112 -4.6 5 10 13 2 1 Se 1

Figure S20. HR-MS of compound **3b**



Figure S22. ¹³C NMR spectrum of **3c**

[Elemental Composition] Page: 1 Data : Dr-Jose-G-Lopez015 Date :04-Mar-2013 17:48 Sample: 577 OOSeNEtOH Note : Luis-Velasco Inlet : Direct Ion Mode : FAB+ Scan#: (11,13) RT : 4.47 min Elements : C 40/0, H 49/0, O 3/0, N 3/0, Se 1/0 Mass Tolerance : 100ppm, 2mmu if m/z >2 Observed m/z Int% 273.9977 100.0 Estimated m/z Error [ppm] U.S.C H O N 273.9982 -2.1 6.5 10 12 3 1 Se 1

Figure S23. HR-MS of compound **3c**



Figure S25. ¹³C NMR spectrum of **3d.**

[Elemental Composition] Data : Dr-Jose-G-Lopez056 Sample: 1412 MeP1SeOH2 Note : Luis-Velasco	Page: 1 Date :23-May-2011 16:17
Inlet : Direct	Ion Mode : FAB+
RT : 0.40 min	Scan#: (6,22)
Elements : C 40/0, H 49/0, O 3/0,	N 3/0, Se 1/0
Mass Tolerance : 100ppm, 2mmu	if $m/z \ge 2$
Observed m/z Int%	
232.0122 7.1	
Estimated m/z Error [ppm] U.S.	C H O N Se
232.0115 3.0 4	8 12 1 2 1

Figure S26. HR-MS of compound 3d



Figure S28. ¹³C NMR spectrum of **3e**

[Elemental Composition] Page: 1
Data : Dr-Jose-G-Lopez016 Date :06-Nov-2012 19:24
Sample: 2772 Caro2
Note : Luis-Velasco
Inlet : Direct Ion Mode : FAB+
RT : 0.31 min Scan#: (6,10)
Elements : C 40/0, H 49/0, O 3/0, N 3/0, Se 1/0
Mass Tolerance : 100ppm, 2mmu if m/z >2
Observed m/z Int%
 282.0285 100.0
Estimated m/z Error [ppm] U.S. C H O N Se
 282.0271 4.9 7 12 14 1 2 1

Figure S29. HR-MS of compound 3e



Figure S31. ¹³C NMR spectrum of **3f**

[Elemental Composition] Page: 1 Date :26-Oct-2012 10:31 Data : Dr-Jose-G-Lopez009 Sample: 2731 SSeNOH Note : Luis-Velasco Ion Mode : FAB+ Inlet : Direct RT : 1.68 min Scan#: (36,43) Elements : C 40/0, H 49/0, O 3/0, N 3/0, S 2/0, Se 1/0 Mass Tolerance : 100ppm, 2mmu if m/z >2 Observed m/z Int% 234.9582 100.0 Estimated m/z Error [ppm] U.S.C H O N S Se 234.9570 4.9 4 7 9 1 1 1 1

Figure S32. HR-MS of compound 3f



[Elemental Composition] Page: 1 Date :26-Oct-2012 10:19 Data : Dr-Jose-G-Lopez008 Sample: 2729 OSeNOH Note : Luis-Velasco Inlet : Direct Ion Mode : FAB+ Scan#: (34,42) RT : 1.64 min Elements : C 40/0, H 49/0, O 3/0, N 3/0, Se 1/0 Mass Tolerance : 100ppm, 2mmu if m/z >2 Observed m/z Int% 218.9785 100.0 Estimated m/z Error [ppm] U.S.C H O N 218.9799 -6.3 4 7 9 2 1 Se 1

Figure S35. HR-MS of compound 3g

8. X-Ray diffraction analyses

Suitable X-ray quality crystals of **3f** were grown by slow evaporation of chloroform at room temperature. A crystal of **3f** was mounted on a glass fiber at room temperature, then placed on a Bruker Smart Apex CCD diffractometer, equipped with Mo KR radiation; decay was negligible in both cases. Details of crystallographic data collected for compound **3f** are provided in Table S2. Systematic absences and intensity statistics were used in space group determination. The structure was solved using direct methods.⁴Anisotropic structure refinements were achieved using full matrix, least-squares technique on all non-hydrogen atoms. All hydrogen atoms were placed in idealized positions, based on hybridization, with isotropic thermal parameters fixed at 1.2 times the value of the attached atom. Structure solutions and refinements were performed using SHELXTL V6.10. ⁵ CCDC-1015044 (**3f**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via <u>www.ccdc.ac.uk/data_request/cif</u>.

	3f
Empirical Formula	C ₇ H ₉ NOSSe
Formula Weight (g mol ⁻¹)	234.17
Crystal size (mm)	0.28 x 0.22 x 0.07
Color	Orange
Crystal system	Monoclinic
Space Group	P21/c
<i>a</i> (Å)	8.595(10)
<i>b</i> (Å)	13.301(15)
<i>c</i> (Å)	8.139(9)
α (°)	90
β (°)	107.864(4)
γ (°)	90
∨ (ų)	885.7(17)
Ζ	4
D _{calc} (g cm ³)	1.756
Number of collected reflections	5879
Number of independent reflections (<i>R</i> int)	2026, <i>R</i> _{int} = 0.0202
Absorption correction method	Semi-empirical from equivalents
Maximum and minimum transmission	0.7456 and 0.5658
Data/restraints/parameters	2026/2/1065
Final <i>R</i> indices $[l>2\sigma(l)]$	<i>R</i> = 0.0292, <i>wR</i> 2 = 0.0703
R índices (all data)	<i>R</i> = 0.0367, <i>wR</i> 2 = 0.0741
Goodness-of-fit on <i>F</i> ²	1.057

9. Biological studies.

Cell lines culture and culture medium.

The compounds **3(a-h)** were screened in vitro against six tumor cell lines: HCT-15 (human colorectal adenocarcinoma), U251 (human glioblastoma), PC-3 (human prostatic adenocarcinoma), MCF-7 (human mammary adenocarcinoma), K562 (human chronic myelogenous leukemia) and SKLU-1 (human lung adenocarcinoma), and the healthy cell line MT2 (Human T-lymphocyte). These cell lines were supplied by National Cancer Institute (USA). The human tumor cytotoxicity was determined using the protein-binding dye Sulforhodamine B (SRB) in microculture assay to measure cell growth, as described in the protocols established by the NCI.²⁶ The cell lines were cultured in RPMI-1640 medium (Sigma Chemical Co., Ltd., St. Louis, MO, U.S.A.) supplemented with 10 % fetal bovine serum which was purchased from Invitrogen Corporation, 2 mM L-glutamine, 10,000 units/mL penicillin G sodium, 10,000 µg/mL streptomycin sulfate and 25 µg/mL amphotericin B (Gibco) and 1 % non-essential amino acids (Gibco). They were maintained at 37°C, in a 5 % CO₂ atmosphere with 95% humidity. The viability of the cells used in the experiments exceeds 95 %, which was determined with trypan blue.

Cytotoxicity assays

The cells were removed from the tissue culture flasks by treatment with trypsin, and then diluted with fresh media. Of this cell suspension, $100 \,\mu$ L containing 5000-10,000 cell per well were pipetted into 96 well microtiter plates (Costar) and the material was incubated for 24 h, at 37 °C in a 5% CO₂ atmosphere.

A 20 mM stock solution of each compound was prepared using ethanol as solvent and the solutions of lower concentrations (50 to 1 μ M) were prepared by accurate dilution. To determine the inhibition of the growth (%) of human tumor cell lines for **3(a-h)** at 50 μ M in EtOH, 100 μ L of an ethanolic solution 50 μ M of each compound were added to each well (Table S2). The final percentage of ethanol in each well was 0.05%. The cultures were exposed for 48 h. After the incubation period, cells were fixed in situ by the addition of 50 mL of cold 50 % (w/v) trichloroacetic acid. The plates were incubated for 1 h, at 4 °C. The supernatant was discarded, and the plates were washed three times with water and air-dried. Cultures fixed with trichloroacetic-acid were stained for 30 min with 100 μ L of 0.4% SRB solution.

Protein-bounded dye was extracted with 10 mM unbuffered tris base and the optical densities were read on a Microplate Reader Synergy HT (Elx 808, BIOTEK Instruments, Inc., U.S.A.), with a test wavelength of 515 nm. Results were expressed as IC_{50} values, they were calculated according to the protocol of Monks,²⁶ were a dose-response curve was plotted for each compound, and the concentration giving 50% inhibition (IC₅₀) was estimated from non-linear regression equations.

A similar procedure was followed for obtaining the IC_{50} of each compound, which was conducted using the three human cancer cell lines that exhibited the best results. For comparative purposes, the MT2 cell line described above was also used. The solutions of test compounds were prepared at concentrations ranging from 1.0 to 50 μ M.

Entry	Compound	HCT-15	U251	PC-3	MCF-7	K562	SKLU-1
1	3a	62.6	39.9	88.3	59.5	25.4	41.8
2	3b	68.7	87.48	100	97.36	46.88	54.3
3	Зc	67.16	71.96	84.14	62.25	90.74	35.66
4	3d	83.92	56.78	74.84	72.15	35.57	21.73
5	3e	>100	91.0	>100	>100	99.0	86.1
6	3f	>100	97.3	>100	78.9	62.15	48.3
7	Зg	62.65	60.49	64.19	51.91	44.62	37.54
8	3h (Lead)	98.51	98.27	88.64	>100	62.68	95.7

Table S3. Inhibition of the growth (%) of human tumor cell lines for 3(a-h) at 50 μM in EtOH

10. - References

- a) E.O. Fischer, A. Maasböl, Angew. Chem. Int. Ed. Engl. 1964, 3, 580; b) J.A. Connor, E.M. Jones, J.P. Lloyd, J. Organomet. Chem. 1970, 24, C20; c) G.M. Chu, I. Fernández, M.A. Sierra, J. Org. Chem. 2013, 78, 865; d) M.L. Lage, I. Fernandez, M.J. Mancheño. M.A. Sierra, Inorg. Chem. 2008, 47, 5253; e) B. van der Westhuizen, P.J. Swarts, L.M. van Jaarsveld, D.C. Liles, U. Siegert, J.C. Swarts, I. Fernández, D.I. Bezuidenhout, Inorg. Chem. 2013, 52, 6674.
- a) J.G. López-Cortés, L.F. Contreras de la Cruz, M.C. Ortega-Alfaro, R.A. Toscano, C. Alvarez-Toledano, H. Rudler, *J. Organomet. Chem.* 2005, 690, 2229; b) J.G. López-Cortés, A. Samano-Galindo, M.C. Ortega-Alfaro, A. Toscano, H. Rudler, A. Parlier, C. Alvarez-Toledano, *J. Organomet. Chem.* 2005, 690, 3664.
- a) A.I. Gutiérrez-Hernández, J.G. López-Cortés, M.C. Ortega-Alfaro, M.T. Ramírez-Apan, J.J. Cázares-Marinero, R.A. Toscano, *J. Med. Chem.* 2012, **55**, 4652. b) A. Ramírez-Gómez, A.I. Gutiérrez-Hernández, M.A. Alvarado-Castillo, R.A. Toscano, M.C. Ortega-Alfaro, J.G. López-Cortés, *J. Organomet. Chem.* 2020 (DOI: 10.1016/j.jorganchem.2020.121315).
- A. Altomare, G. Cascarano, C. Giacovazzo, A. Guagliardi, M.C. Burla, G.Polidori, M. Canalli, *J. Appl. Crystallogr.* 1994, 27, 435-436.
- 5) G.M. Sheldrick, Acta Crystallogr. 2008, A64, 112-122.